A pH-responsive polypseudorotaxane has been synthesized via cucurbit[6]uril (CB6)-catalyzed 1,3-dipolar cycloaddition using diazide and diaclyne monomers, which contain a long aliphatic spacer. The polypseudorotaxane was characterized by spectroscopic techniques (1H, 13C NMR and FT-IR) and matrix assisted laser desorption time-of-flight mass spectrometry (MALDI-TOF MS). The experimental results reveal that this polypseudorotaxane behaves as a pH-driven polymeric switch. Thus, when amine groups are protonated at an appropriate pH, CB6s are located on the triazole rings due to ion–dipole interaction, whereas at high pH they move onto the hydrophobic aliphatic spacer rather than slipping off the polypseudorotaxane.

**Result and discussion**

**Synthesis and characterization of monomers and polypseudorotaxane**

Monomers dialkyne 2 and diazide 3 and polypseudorotaxanes 4 were prepared as shown in Scheme 1. The monomer 2 was synthesized by the reaction of dibromododecane with excess propargylamine in chloroform followed by the protonation of the dipropargylic amine species using 0.1M HCl solution in diethylether. The resulting hydrochloride salt was purified by recystallization from water to produce 2 in 75% yield.

To synthesize monomer 3, first chloroethylamine hydrochloride was converted to azidoethylamine by a nucleophilic substitution reaction with sodium azide in water. Then, 1,12-dibromododecane was N-alkylated by treatment with excess azidoethylamine followed by the conversion of the diazidoamine species to its hydrochloride salt 3. The monomer 3 was obtained in 80% yield after recystallization from ethanol.
The monomers 2 and 3 were characterized by spectroscopic methods (\(^1\)H and \(^{13}\)C NMR, FT-IR), as well as elemental analysis. The results are in good agreement with the proposed structures.

To synthesize the polypseudorotaxane 4, one equiv. of monomer 2 and one equiv. of monomer 3 were dissolved in water at 60 °C followed by the addition of two equiv. of CB6 in 6M HCl. The resulting solution was stirred at 60 °C and after 48 h the solvent was removed under reduced pressure. The resulting light yellow film was redissolved in water and purified by dialysis using a membrane with a nominal molecular weight cut-off of 5 kDa. Polypseudorotaxane was obtained in 80% yield as a colourless film after the removal of solvent and drying the film under vacuum at 40 °C for 12 hr.

The formation of polypseudorotaxane 4 was confirmed first by \(^1\)H NMR spectroscopy. Fig. 1 compares the \(^1\)H NMR spectra of the monomers 2 and 3, and the polypseudorotaxane 4.

Upon the formation of 4, a new resonance at 6.5 ppm appears which is a diagnostic signal for the proton of a threaded triazole ring (Fig. 1c). In addition, the signal of the terminal proton of alkyn protons at 2.9 ppm is not observed which is a diagnostic signal for the proton of a dodecamethylene spacer.

One can also determine the location of CB6 from the \(^1\)H-NMR spectrum of 4. If the dodecamethylene spacer was threaded by CB6, the resonances due to its protons would shift significantly to upfield because of the shielding effect of CB6. However, the spectrum clearly shows that CB6 is localized on the triazole ring, but not on the dodecamethylene spacer.

The number-average molecular weight of the polypseudorotaxane can be estimated by comparing the integral intensities of the threaded triazole proton (\(t\)) to the central methylene protons of dodecamethylene spacer (\(C\) and \(D\)) at 6.5 and 1.5 ppm respectively (Scheme 1). Each repeat unit, which has the molecular weight of approximately 2600 Da, contains two triazole protons (\(t\)) and a total of 32 central methylene protons (\(C\) and \(D\)). The ratio of \(t\) to \(C+D\) should be 1 : 16 for an infinite polymer chain. From the integration, the ratio was calculated as 1 : 17 (\(t/(C+D)\)) which corresponds to repeat unit of eight. Thus, the number-average molecular weight of the polymer corresponds to about 20800 Da.

In the IR spectrum of 4, a very weak signal due to azide stretching at 2200 cm\(^{-1}\) was observed, which probably belongs to an azide end group rather than to the unreacted monomer. Having very weak signal for the azide stretching also indicates the formation of a polypseudorotaxane, because when we mix mechanically one equivalent of dialkyne and one equivalent of diazide monomers, and two equivalents of CB6 and then record the IR spectrum of this mixture, we observe the azide stretching peak with a very strong intensity.

It is known that the molecular weight determination of electrolytes like ours is quite troublesome. So we were not able to determine the molecular weight of the polypseudorotaxane with techniques other than the matrix assisted laser desorption time-of-flight mass spectrometry (MALDI-TOF MS).

MALDI-TOF MS verifies the formation of 4 (Fig. 2). The positive ion MALDI-MS spectrum of polypseudorotaxane was recorded using 3-indoleacrylic acid as a matrix. Peak groups were observed repeating by approximately 2600 Da mass unit intervals. Beside the potassium adducts of dimer, tetramer, hexamer, octamer, decamer and dodecamer other unidentified peaks were detected. Maximum mass was observed around 15600 Da which corresponds to about six repeating units, that is basically 12 CB6s threaded triazoles.

In the MALDI MS spectrum, we would not expect to observe the molecular ions lower than 5000 Da because the polymer was purified by dialysis using a membrane with a
nominal molecular weight cut-off of 5 kDa for a prolonged time. Nevertheless, low molecular weight ions with high intensity in MS spectrum were detected. One can speculate that the dialysis may cause the dethreading of CB6 macrocycles from the polymer and consequently leave behind a low molecular weight polymer. This possibility can be ruled out because the 1H-NMR spectrum of the dialysed poly-pseudorotaxane does not reveal the presence of chemical shifts due to unencapsulated triazole protons. Moreover, the ratio of threaded triazole proton to central methylene protons of dodecyl spacer even increased after the dialysis. Thus, having intense peaks for low molecular weight ions can be attributed to the fact that MALDI MS does not provide quantitative information on the molecular weight distribution for polyelectrolytes because the polymer chains with low molecular weight are transferred to gas phase more easily. Low mass signals might arise from fragmentation of the poly-pseudorotaxane under laser energy or due to dethreading from the pseudorotaxane. Upon protonation of the nitrogens, it moves back on the triazole ring, where via ion–dipole interaction it forms stronger complexes with each of the two positively charged nitrogen atoms on the amines (Scheme 2).

Investigation of pH-driven switching properties of poly-pseudorotaxane 4

Next, we attempted to dethread CB6 from poly-pseudorotaxane 4 to obtain the polytriazole. In order to do this, the ion–dipole interactions between the diammonium ions and carbonyl groups of CB6 must first be disrupted by deprotonation of the ammonium ions with a base.

To perform the dethreading experiment, 4 was dissolved in water and 0.1M aqueous solution of NaOH was added to this solution. Instantly a white gel formed; it was isolated by centrifugation and washed with water several times and then with 0.2 M aqueous solution of NaCl to remove CB6, in case any was left. The wash solution was evaporated to dryness and the IR spectrum of the residue was recorded, however, no CB6 was detected.

The isolated solid was found to be insoluble in any common organic solvent. However, when it was treated with 0.1M HCl solution, it became soluble. Interestingly, it was found that the 1H-NMR spectrum of this solid (in D2O–HCl) and its IR spectrum are similar to the spectrum of poly-pseudorotaxane 4.

This result suggests that once the ion–dipole interaction between the ammonium ions and the carbonyl groups of CB6 is disrupted at an appropriate pH, CB6 shuttles to the more hydrophobic part, the dodecamethylene spacer, rather than dethreading from the pseudorotaxane. Upon protonation of the nitrogens, it moves back on the triazole ring, where via ion–dipole interaction it forms stronger complexes with each of the two positively charged nitrogen atoms on the amines (Scheme 2).

To reveal the details of the pH-driven movement of CB6 in 4, we followed the acid–base switching by 1H NMR spectroscopy; the spectra of 4 in D2O were recorded at varying pHs using NaOH and HCl (Fig. 3). They were recorded in each case about five minutes later addition of base or acid to the solution of poly-pseudorotaxane.

Up to pH 9, no significant spectral changes were observed. As shown in Fig. 3b, at pH 9, the intensity of the resonance due to the encapsulated triazole proton at 6.5 ppm decreases, whereas three weak signals around at 8 ppm due to unencapsulated triazole ring proton emerge. The appearance of three different signals due to unencapsulated triazole proton indicates that they are in different chemical environments. Furthermore, a new signal at 2.5 ppm appears; this signal can be assigned to protons b by assuming that they are adjacent to the carbonyl groups of CB6 and as a result, shifted downfield. All these spectral changes can be ascribed to the partial movement of CB6 rings from the triazole site to the dodecamethylene spacer that driven by the partial deprotonation of ammonium ions at pH 9.

The CB6 rings are probably quite mobile and moving back and forth over the long dodecamethylene spacer. As a result, averaged signals due to the protons of central methylene
groups of dodecamethylene spacer are observed in the 1H-NMR spectrum. At pH 9, from the integration of resonances due to threaded triazole proton and CB6 protons, it is estimated that approximately 50% of CB6s are located on the triazole part and the rest on the dodecamethylene spacer, whereas at pH 10 (Fig. 3c), approximately 20% of CB6 reside on the triazole part and the rest on the dodecamethylene part. At pH 11, the 1H NMR spectrum reveals that almost 95% of CB6s reside on the dodecamethylene spacer. Increasing the pH of the solution beyond 11 causes immediate precipitation. However, the addition of HCl causes the dissolution of the precipitate in a few minutes. 1H-NMR spectrum is recorded about five minutes later the addition of HCl (pH \(y\) 5) to the precipitate. The spectrum reveals that most of the CB6s shuttle back to their initial location, which is the triazole site. The signal due to encapsulated proton at 6.5 ppm reappears and the signal at 2.5 ppm almost disappears.

Conclusions

In this paper, we reported the synthesis and characterization of a novel pH-responsive CB6-based polypseudorotaxane prepared via 1,3-dipolar cycloaddition catalyzed by CB6. The result confirms the critical role of spacers employed in the monomers for catalytic self-threading synthesis of polypseudorotaxanes.

The experimental results reveal that the polypseudorotaxane behaves as a pH-driven polymeric switch. Thus, once the ion–dipole interaction between the ammonium ions and the carbonyl groups of CB6 is disrupted at an appropriate pH, CB6 shuttles to the hydrophobic part of the polymer, the dodecamethylene spacer, rather than de-threading from the polypseudorotaxane. Hence, it forms a relatively stable complex with the dodecamethylene spacer via hydrophobic interaction. However, upon protonation of the amine groups, it moves back on the triazole ring, where via ion–dipole interaction it forms stronger complexes with each of the two positively charged nitrogen atoms on the amines.

A detailed investigation of the properties of this novel polypseudorotaxane and the synthesis of its polytriazole by de-threading, and using other approaches are currently in progress.

Experimental

CB6 was prepared according to literature. All other reagents and solvents were of the commercial reagent grade and used without further purification except where noted. Nuclear magnetic resonance (NMR) spectra were recorded on a Bruker Avance DPX-400 MHz spectrometer. In all cases, samples were dissolved in D_2O with DSS (3-(trimethylsilyl)-1-propanesulfonic acid sodium salt) as an external standard.

Mass spectra were acquired on a Voyager-DETM PRO MALDI-TOF mass spectrometer (Applied Biosystems, USA) equipped with a nitrogen UV-Laser (\(\lambda = 337\) nm) at 10-7 Torr and delayed extraction mode (delay 300 ns) by changing the laser intensity, if necessary. All spectra were acquired in reflector mode using 20 kV acceleration potential and they were the average of 200 laser shots. Then, data were transferred to a personal computer for further processing.

3-Indoleacrylic acid (IAA) (Fluka, Switzerland) was chosen as the best MALDI matrix and prepared in MeOH at a concentration of 20 mg mL\(^{-1}\). MALDI samples were prepared by mixing the sample solution with the IAA matrix solution (1 : 10 v/v) in a 0.5 mL Eppendorf\(\text{®}\) micro tube. Finally 0.1 mL of these mixtures were deposited on the sample plate, dried in room temperature and then analyzed.

Dialysis was carried out in an Amicon8200 ultrafiltration stirred cell (200 mL) and polysulfone membranes (NMWCO 5 kDa). Approximately 2 L water was eluted through the ultrafiltration cell under 4.5 bar nitrogen pressure.

Synthesis of \(N,N’\)-di-prop-2-ynyldodecane-1,12-diamine dihydrochloride 2

A solution of 1,12-dibromododecane (0.93 g, 2.8 mmol) in CHCl\(_3\) (5 mL) was added dropwise to excess propargylamine...
(1.56 g, 28.4 mmol) at room temperature. The resulting solution was stirred at room temperature for 48 h. 0.1M Aqueous NaOH (5 mL) was added to reaction mixture and the mixture was stirred for 1 h at room temperature. The organic layer was separated by extraction and washed with saturated solution of NaCl. After drying the organic layer over CaCl₂, the solvent was removed under reduced pressure. A light yellow solid residue was obtained. It was dissolved in diethyl ether (5 mL) and the solution was cooled to 0°C in an ice-bath. 0.1 M HCl in diethyl ether (5 mL) was added dropwise. The resulting mixture was stirred at room temperature for 4 h. A beige precipitate was collected and recrystallized from water to obtain off-white solid.

Yield: 0.75 g, (75%). M.p.: 218–219°C

Elemental Analysis for C₁₈H₃₄Cl₂N₂
Calc.: C, 61.88; H, 9.81; Cl, 20.29; N, 8.02; Found: C, 61.69; H, 9.35; N, 7.90

IR (KBr disk, cm⁻¹): 3480 (w), 3385 (m), 3135 (m), 2452 (m), 2200 (w).

1H NMR (400 MHz, D₂O): δ 1.3 (m, 16H, e, f), 1.6 (m, 4H, d), 2.9 (t, 2H, 4JHH 2.2 Hz, b), 3.1 (t, 4H, 3JHH 7.8 Hz, c), 3.9 (d, 4H, 4JHH 2.2 Hz, a).

13C NMR (100 MHz, D₂O): δ 25.3 (i), 25.6 (h), 28.2 (g), 28.4 (f), 28.6 (e), 36.2 (c), 46.8 (d), 73.2 (b), 77.9 (a).

Synthesis of N,N'-bis(2-azido-ethyl)dodecane-1,12-diamine dihydrochloride 3

A solution of 1,12-dibromododecane (0.86 g, 2.6 mmol) in CHCl₃ (5 mL) was added dropwise to freshly prepared azidoethylamine (2.25 g, 26.2 mmol) at room temperature. The resulting solution was stirred at room temperature for 48 h. 1M aqueous NaOH (5 mL) was added to reaction mixture and the mixture was stirred for 1 h at room temperature. The organic layer was separated by extraction and washed with saturated NaCl. After drying the organic layer over CaCl₂, the solvent was removed under reduced pressure. A light yellow solid residue was obtained. It was dissolved in diethyl ether (5 mL) and the solution was cooled to 0°C in an ice-bath. 0.1 M HCl in diethyl ether (5 mL) was added dropwise. The resulting mixture was stirred at room temperature for 4 h. The solvent was removed under reduced pressure and off-white solid residue was recrystallized from ethanol.

Yield: 0.89 g, (80%). M.p.: > 208°C (decomposed)
References


